

Effect of microbes on Patterns of Labeled White Blood Cells in Osteomyelitis

Saroj Kumar Sahu¹, Sudatta Ray²

^{1,2}Associate Professor, Department of Nuclear Medicine, IMS and SUM Hospital, Siksha 'O' Anusandhan Deemed to be University, Bhubaneswar, Odisha, India.

How to cite this article: Saroj Kumar Sahu, Sudatta Ray. Effect of microbes on Patterns of Labeled White Blood Cells in Osteomyelitis. Indian Journal of Public Health Research and Development 2023;14(3).

Abstract

Aim: This study investigates the effect of microbiological characteristics of causative organisms on the scintigraphic patterns of labelled white blood cell (WBC) scans in cases of proven osteomyelitis.

Methods: Retrospective analysis of 25 patients referred with suspected osteomyelitis and had both bone and labelled WBC scans performed and complete records of the microbiological culture of the causative organism. The bone and labelled WBC scans were retrieved and reviewed by two nuclear medicine physicians. Any definite focal accumulation of labelled WBCs within the bone was considered positive for osteomyelitis. Diagnosis of osteomyelitis in the discharge summary was considered the reference standard and was based on a combination of the clinical scenario, imaging, and laboratory findings including microbiology. A correlation of the pattern of labelled WBC and the type of microorganisms was done.

Results: A total of 16 patients were included in this study, seven females and nine males. Of these, seven patients had Gram-positive whereas nine patients had Gram-negative organisms. The majority (85.7%) of Gram-positive organisms showed increased accumulation of labelled WBCs, whereas only one-third (33.3%) of patients with Gram-negative organisms had such findings.

Conclusion: The pattern observed in this study shows that the falsenegative results of labelled WBC scans were mainly noted in patients with Gram-negative as opposed to Gram-positive infections. This confirms the experimental animal study findings that the secretion of anti-chemotactic factors by Gram-negative organisms, seems to be inhibiting the migration of labelled WBCs to the site of infection. The inhabitation is decreasing the accumulation of labelled WBCs and consequently resulting in a falsenegative finding. The study adds to evidence that microbiological characteristics of the causative organisms are another explanation for the falsenegative WBC in proven osteomyelitis.

Keywords: Bone scan, Gram-negative bacteria, Gram-positive bacteria, labelled white blood cells, osteomyelitis.

Introduction

The diagnosis of osteomyelitis can be challenging and depends on clinical findings, laboratory tests, and

imaging studies supported by tissue culture. Gram-positive bacteria are the primary causative agents of osteomyelitis and *Staphylococcus aureus* is by far the most common in all age groups.¹ Osteomyelitis can

Corresponding Author: Saroj Kumar Sahu, Associate Professor, Department of Nuclear Medicine, IMS and SUM Hospital, Siksha 'O' Anusandhan Deemed to be University, Bhubaneswar, Odisha, India.

E-mail: drsarojkumarsahoo@gmail.com

also be caused by Gram-negative bacteria, especially in patients with certain predispositions such as sickle cell disease, diabetes, and trauma.² Multiple morphologic and functional imaging modalities have been used with different sensitivities and specificities. Conventional radiographs are usually used first due to their wide availability and low cost. However, plain radiographs usually have limited value due to their low sensitivity for acute infections as the anatomical changes are usually seen in the late stages of the disease.³ Computed tomography (CT) provides good anatomical information. However, its ability to differentiate bone from soft tissue infection is limited. In addition, its value is diminished in postsurgical cases with a metallic implant.³ On the other hand, magnetic resonance imaging (MRI) provides high-resolution images with a good ability to differentiate soft tissue from a bone infection.⁴ It has high sensitivity in detecting osteomyelitis, but its specificity is limited by many causes of false positive results such as postsurgical changes, bony infarction, and Charcot's joint.⁵ Multiple nuclear medicine procedures are used in diagnosing osteomyelitis. Multi-phase bone scintigraphy using technetium-99m diphosphonates (Tc-99m) provides an early diagnosis with comparable sensitivity to MRI. However, it has low specificity as multiple potential causes can result in false positive results.⁶ The specificity of a bone scan can be improved when it is combined with a labelled white blood cell (WBC) scan. The labelled WBC study is the most accurate in diagnosing acute osteomyelitis. The combined approach using bone and labelled WBC scans provide comparable sensitivity and superior specificity in comparison to MRI.⁷ Although the combined approach has generally acceptable accuracy (ranging from 79% to 100%), many factors should be considered before choosing this approach. The duration (acute or chronic) and the location (axial or appendicular) of the infection may affect the accuracy of this technique.⁸ Furthermore, the microbiological characteristics of the causative organisms may also influence the accuracy of the procedure.

Multiple studies reported low sensitivity of labelled WBC scans for the detection of vertebral osteomyelitis. Palestro *et al.* reported a low sensitivity (39%) of In-111 WBC in 28 patients with proven vertebral osteomyelitis.⁹ Another study on 22 patients

with spine infections reported a sensitivity of only 17%.¹⁰ Hovi also reported three cases of histologically proven osteomyelitis detected by MRI but none by Tc-99m-hexamethyl-propylene-amine oxime (99mTc-HMPAO)-labelled WBC studies.¹¹

The exact reason for this observation has been uncertain, and there are several possible explanations. The diagnosis of vertebral osteomyelitis is often delayed, and most infections become chronic which lowers the sensitivity of labelled WBC scans.¹² Another possible explanation is that infection results in very high pressure in the vertebra and therefore, labelled WBCs could not migrate to the focus of infection during the available time for imaging. Palestro and Love proposed that marrow uptake of labelled WBCs in the normal spine may be greater than in the infected bone, masking the abnormality or causing a relative photogenic region.¹³ Finally, Fernandez-Ulloa *et al.* suggested that Gram-negative organisms could be behind this finding since they secrete anti-chemotactic factors.¹⁴ A preliminary animal study with rabbits confirmed that Gram-negative infections in vertebral and femoral osteomyelitis are associated with low to no accumulation of labelled WBCs.¹² This study aims to investigate the microbiological characteristics of labelled WBC scans in cases of nonvertebral osteomyelitis since the high-pressure theory is not a factor in such sites.

Materials and Methods

Retrospective review of cases referred to the Department of Nuclear Medicine for suspected osteomyelitis from September 2009 to September 2013. Only 25 patients, who had both bone and labelled WBC scans performed were diabetics, and only two out of the seven patients with Gram-positive infections are diabetics (Table 1). Seven patients had Gram-positive bacterial infection, whereas nine had Gram-negative infection (Table 2). Out of the seven cases with Gram-positive infection, six (85.7%) had increased uptake of labelled WBCs [Table 3]. The majority of the positive labelled WBC scan cases of Gram-positive bacterial infection were due to *pus* (83.3%). Out of the nine Gram-negative bacterial-infected cases (Table 4), only three had increased uptake (33.3%), whereas the other six had no uptake (66.7%) on their labelled WBC scans. Two of the three patients with increased

labelled WBCs uptake were infected with *Enterobacter aerogenes* while the third patient was infected with multiple Gram-negative organisms (Table 3). Figure 1 illustrates an example of Gram-negative bacterial infection at the foot with nouptake of labelled WBCs and complete records of the microbiological culture of the causative organism, were included in the study. The data were collected and attained from the nuclear medicine Department picture archiving and communication system, patients' hospital files and Laboratory Information System. Data from these selected 25 patients with complete records were analyzed. The three-phase bone scan was performed following the intravenous administration of 740-1,110 MBq (20-30 mCi) of Tc-99m-methylene diphosphonate. The dynamic blood flow images were acquired immediately after the radiotracer administration over the region of interest. The blood pool images were obtained after 5 min. Delayed whole-body images, as well as spot images, were obtained after 3 h. Single-photon emission CT (SPECT) and SPECT/CT images were also obtained on selected cases when an accurate anatomical correlation was needed.

Labeled-WBC scan was performed the following day. The patient WBCs were labelled with 25 mCi of ^{99m}Tc-HMPAO and re-injected intravenously. Whole-body and spot images of the area of interest were acquired at 2 and 4 h after the radiotracer administration. Furthermore, SPECT and SPECT/CT images were acquired on selected patients for correlation with the bone scan. The bone and labelled WBC scans were retrieved and reviewed by two nuclear medicine physicians. The images were interpreted based on a visual assessment of the uptake pattern of labelled WBC in correlation to the bone scan. Any definite focal accumulation of labelled WBCs within the bone was considered positive for osteomyelitis. All patients selected in the analysis had also microbiological and inflammatory markers performed. The final diagnosis of osteomyelitis in the discharge summary was considered the reference standard and was based on a combination of the clinical scenario, imaging, and laboratory findings including microbiology.

Results

Among the 25 patients studied, seven patients were excluded from further analysis since osteomyelitis was not their final diagnosis. In addition, two patients with proven osteomyelitis were excluded from the final analysis as they had a combination of mixed Gram-positive and Gram-negative bacteria in the culture. The remaining 16 patients including seven females and nine males with ages ranging between 20 and 75 years (mean 51.2) were included in the final analysis. None of the patients was given antibiotics for >48 h before scans. In the investigated cases, the osteomyelitis was located at the tibia in five cases, three cases at the metatarsal, two at each of the talus, femur, and calcareous, as well as one case at each of the radius and posterior talocalcaneal joint. Eight out of the 16 patients included were diabetics. The majority of the patients with Gram-negative infections (six out of nine, 66.7%) were diabetics, and only two out of the seven patients with Gram-positive infections are diabetics (Table 1). Seven patients had Gram-positive bacterial infection, whereas nine had Gram-negative infection (Table 2). Out of the seven cases with Gram-positive infection, six (85.7%) had increased uptake of labelled WBCs (Table 3). The majority of the positive labelled WBC scan cases of Gram-positive bacterial infection were due to *eus* (83.3%).

Table 1: Demographics Data of patients

No. of Patient	Scan	Gram stain	Site of infection	Diabetic
1	+	+	Talus	-
2	-	-	Metatarsal	+
3	+	+	Femur	-
4	-	-	Tibia	+
5	+	-	Tibia	+
6	+	+	Tibia	-
7	-	-	Talus	-
8	-	-	Tibia	-
9	-	-	Metatarsal	-
10	+	+	Calcaneus	+
11	+	+	Calcaneus	+
12	-	-	Metatarsal	+

Continue....

13	+	-	Posterior talocalcaneal Joint	+
14	-	+	Radius	-
15	+	+	Femur	-
16	+	-	Tibia	+

Table 2: Type of micro-organisms and labelled white blood cells scan findings

Organism	Number of cases	Positivescan (%)	Negativescan (%)
Gram-positive	8	6(85.7)	2(14.3)
Gram-negative	7	3(33.3)	4(66.7)

Table 3: Results of labelled white blood cell scan in patients with Gram-positive bacterial infections

Patient number	Scan results	Organism
1	+	Staphylococcus aureus
2	+	Enterococcus faecalis
3	+	Staphylococcus aureus
4	+	Enterococcus faecalis
5	+	Staphylococcus aureus
6	+	Enterococcus faecalis
7	+	Staphylococcus aureus

Table 4: Results of labelled white blood cell scan in patients with Gram-negative bacterial infections

Patient number	Scan results	Organism
8	-	Morganella morgagnii <i>Bacterioids spp.</i>
9	-	<i>Klebsiella pneumoniae</i>
10	-	Enterobacter sakazakii
11	-	Enterobacter aerogenes Morganella morgagnii
12	+	Escherichia coli Pseudomonas spp. Bacterioids fragalis
13	+	Enterobacter aerogenes
14	-	Enterobacter aerogenes
15	+	Enterobacter aerogenes
16	-	Citrobacter koseri

Out of the nine Gram-negative bacterial-infected cases (Table 4), only three had increased uptake (33.3%), whereas the other six had noupake (66.7%) on their labelledWBC scans. Two of the three patients with increased labelled WBCs uptake were infected with *Enterobacter aerogenes* while the third patient was infected with multiple Gram-negative organisms (Table 3). Figure 1 illustrates an example of Gram-negative bacterial infection at the foot with noupake of labelled WBCs.

Discussion

Gram-positive organisms are known to be the main cause of osteomyelitis; however, Gram-negative organisms such as *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* can also cause osteomyelitis, especially in the diabetic foot, hospital-acquired infections, and intravenous drug abusers.^{15,16} In this study, seven of the 16 patients with proven osteomyelitis were infected with Gram-positive organisms while nine patients had Gram-negative bacterial infections. The majority of patients (85.7%) with Gram-positive infections had positive labelled WBC scans. As well, the majority of patients (66.7%) with Gram-negative infections had negative labelled WBC scans. The majority of patients in this study were infected with Gram-negative organisms which were likely due to the higher presence of diabetes (66.7%) in this group. However, the main goal of this study was to investigate the pattern of labelled WBC uptake in Gram-negative-caused osteomyelitis. The pattern observed in our small group of patients with nonvertebral osteomyelitis is consistent with the findings advocated by Fernandez-Ulloa *et al.* in their five cases of vertebral osteomyelitis. They noted that the abnormally decreased uptake of labelled WBCs in patients with proven vertebral osteomyelitis was mainly observed in the presence of Gram-negative organisms.¹⁴ Kim *et al.* in 1987¹⁷ and Palestro *et al.* in 1991⁹ have also reported false-negative labelled WBC scans in patients with proven osteomyelitis mainly in cases associated with Gram-negative infections. It has been proposed that false-negative labelled WBC scans of vertebral osteomyelitis are likely due to increased compartmental pressure which may prevent the accumulation of labelled WBCs.¹⁴ However, observing the same trend of decreased accumulation of labelled WBCs in nonvertebral osteomyelitis in animal and human studies indicates.

Conclusion

The pattern observed in this study shows that the falsenegative results of labelled WBC scans were mainly noted in patients with Gram-negative as opposed to Gram-positive infections. This confirms the experimental animal study findings that the secretion of anti-chemotactic factors by Gram-negative organisms, seems to be inhibiting the migration of labelled WBCs to the site of infection. The inhabitation is decreasing the accumulation of labelled WBCs and consequently resulting in a falsenegative finding. The study adds to evidence that microbiological characteristics of the causative organisms are another explanation for the falsenegative WBC in proven osteomyelitis.

Ethical statement: This study was approved institutional ethical committee

Conflict of Interests: The authors declare that they have no conflicts of interest

Source of Funding: Self-financed

References

1. Kumar JV, Robbins SL. Acute and chronic inflammation. In: Cotran RS, Kumar JV, Collins T, Robbins SL, editors. Robbins Pathologic Basis of Disease. 6th ed. Philadelphia: SaundersElsevier; 1999. p. 25-46.
2. Carek PJ, Dickerson LM, Sack JL. Diagnosis and management of osteomyelitis. *Am Fam Physician* 2001;63:2413-20.
3. Kaim AH, Gross T, von Schulthess GK. Imaging of chronic posttraumatic osteomyelitis. *Eur Radiol* 2002;12:1193-202.
4. Kruskal JB. Can USPIO-enhanced spinal MR imaging help distinguish acute infectious osteomyelitis from chronic infectious and inflammatory processes? *Radiology* 2008;248:1-3.
5. Gold RH, Tong DJ, Crim JR, Seeger LL. Imaging the diabetic foot. *Skeletal Radiol* 1995;24:563-71.
6. Hatzenbuehler J, Pulling TJ. Diagnosis and management of osteomyelitis. *Am Fam Physician* 2011;84:1027-33.
7. Ivancević V, Dodig D, Livaković M, Hancević J, Ivancević D. Comparison of three-phase bone scan, three-phase 99m-Tc-HM-PAO leukocyte scan and 67-gallium scan in chronic bone infection. *Prog Clin Biol Res* 1990;355:189-98.
8. de Winter F, van de Wiele C, Vogelaers D, de Smet K, Verdonk R, Dierckx RA, *et al.* Fluorine-18 fluorodeoxyglucose-position emission tomography: A highly accurate imaging modality for the diagnosis of chronic musculoskeletal infections. *J Bone Joint Surg Am* 2001;83-A: 651-60.
9. Palestro CJ, Kim CK, Swyer AJ, Vallabhajosula S, Goldsmith SJ. Radionuclide diagnosis of vertebral osteomyelitis: Indium-111-leukocyte and technetium-99m-methylenediphosphonate bone scintigraphy. *J Nucl Med* 1991;32:1861-5.
10. Whalen JL, Brown ML, McLeod R, Fitzgerald RH Jr. Limitations of indium leukocyte imaging for the diagnosis of spine infections. *Spine (Phila Pa)* 1976;1991;16:193-7.
11. Hovi I. Complicated bone and soft-tissue infections. Imaging with 0.1 T MR and 99mTc-HMPAO-labeled leukocytes. *Acta Radiol* 1996;37:870-6.
12. Elgazzar AH, Dannoon S, Sarikaya I, Farghali M, Junaid TA. Scintigraphic patterns of indium-111 oxine-labelled white blood cell imaging of gram-negative versus gram-positive vertebral osteomyelitis. *Med Princ Pract* 2017;26:415-20.
13. Palestro CJ, Love C. Radionuclide imaging of musculoskeletal infection: Conventional agents. *Semin Musculoskelet Radiol* 2007;11:335-52.
14. Fernandez-Ulloa M, Vasavada PJ, Hanslits ML, Volarich DT, Elgazzar AH. Diagnosis of vertebral osteomyelitis: Clinical, radiological and scintigraphic features. *Orthopedics* 1985;8:1144-50
15. Waldvogel FA, Medoff G, Swartz MN. Osteomyelitis: A review of clinical features, therapeutic considerations and unusual aspects. *N Engl J Med* 1970;282:198-206.
16. Kahn DS, Pritzker KP. The pathophysiology of bone infection. *Clin Orthop Relat Res* 1973;96:12-9.
17. Kim EE, Pjura GA, Lowry PA, Gobuty AH, Traina JF. Osteomyelitis complicating fracture: Pitfalls of 111In leukocyte scintigraphy. *AJR Am J Roentgenol* 1987;148:927-30.
18. Kumar JV, Abbas AK, Astor JC, editors. Inflammation and repair. In: Robbins and Cotran Pathologic Basis of Disease. 9th ed. Philadelphia: Elsevier-Saunders; 2015. p. 69-112.
19. Torda AJ, Gottlieb T, Bradbury R. Pyogenic vertebral osteomyelitis: Analysis of 20 cases and review. *Clin Infect Dis* 1995;20:320-8.
20. Song KS, Ogden JA, Ganey T, Guidera KJ. Contiguous discitis and osteomyelitis in children. *J Pediatr Orthop* 1997;17:470-7.